

The effect of remifentanil on respiratory control, assessed from resting physiological variability

Georgios D. Mitsis, *Member, IEEE*, Ricardo J.M. Governo, Richard Rogers and Kyle T.S. Pattinson

Abstract — We evaluated the influence of a steady state infusion of a model opioid, remifentanil, on respiratory variability during spontaneous respiration in a group of 11 healthy human volunteers. We used linear and nonlinear dynamic models to examine the effects of remifentanil upon both directions of the ventilatory loop, i.e. on the influence of natural variations in end-tidal carbon dioxide (PETCO₂) on ventilatory variability, and vice versa. Breath-by-breath recordings of expired CO₂ and respiration were made, for 15 minutes at estimated effect site (i.e. brain tissue) concentrations of 0, 0.7, 1.1, and 1.5 ng/ml respectively. The results revealed a decrease in the strength of the dynamic effect of PETCO₂ variability on V_T and an increase in the effect of V_T variability upon PETCO₂. Nonlinear models explained these dynamic interrelationships better than linear models. Our approach allows detailed investigation of drug effects in the resting state using non-invasive and minimally perturbing experimental protocols, which can closely represent real life clinical situations.

I. INTRODUCTION

Respiratory depression is the most common serious side effect of opioid drugs [1]. Opioids are prescribed to millions of patients around the world every day, often in unmonitored environments; therefore, avoiding respiratory depression remains an important clinical aim. Opioids depress chemosensitive and rhythm generating centers in the brainstem [2], leading to slowing and increasing irregularity of the respiratory rhythm [3].

Much of the understanding of human respiratory control is based upon characterization of the ventilatory feedback loop, which is shown in Fig. 1 in a simplified form, supplemented by inferences from work in animals.

Manuscript received April 15, 2009. This work was supported in part by the European Social Fund (75%) and National Resources (25%) - Operational Program Competitiveness - General Secretariat for Research and Development (Program ENTER 04), by the Association of Anaesthetists of Great Britain and Ireland and the International Anesthesia Research Society.

Georgios D Mitsis was with the Institute of Communications and Computer Systems, National Technical University of Athens, Athens, Greece and the Nuffield Department of Anaesthetics, University of Oxford, Oxford, UK. He is now with the Department of Electrical and Computer Engineering, University of Cyprus, Nicosia 1678, Cyprus (phone: +357-22892239; fax: +357-22892260; e-mail: gmitsis@ucy.ac.cy).

Ricardo J. Governo is with the Brighton and Sussex Medical School, University of Sussex, Brighton, East Sussex, UK.

Richard Rogers is with the Nuffield Department of Anaesthetics, University of Oxford, Oxford, UK.

Kyle T.S. Pattinson is with the Nuffield Department of Anaesthetics, University of Oxford, Oxford, UK and the Oxford Centre for Functional Magnetic Resonance Imaging of the Brain, University of Oxford, Oxford, UK.

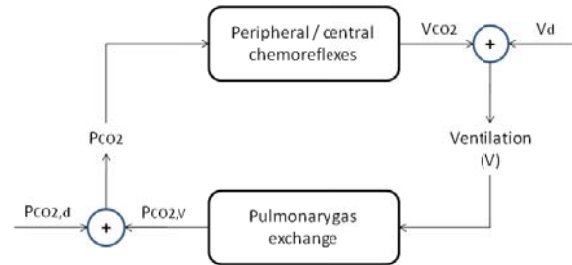


Fig. 1. Simplified diagram of the ventilatory feedback loop. Spontaneous ventilatory variability arises from a chemical component V_{CO_2} that is due to variations in arterial CO₂ tension variability P_{CO_2} , and a non-chemical component V_d (disturbance term, which includes sighs) that is due to all other physiological influences. Similarly, spontaneous P_{CO_2} variability arises from a ventilatory related component $P_{CO_2,V}$ that is due to variations in V_T and a non-ventilatory related component $P_{CO_2,d}$.

Ventilatory responses are usually examined during hypoxic or hypercapnic stimulation [4]; specifically, depression of these responses by opioids has been well reported [5][6]. Moreover, it has been shown that spontaneous breath-to-breath fluctuations in end-tidal CO₂ (PETCO₂) are responsible for a considerable fraction of the normal variability in tidal volume (V_T) [7], and the dynamic effects of these spontaneous fluctuations have been used to derive information on ventilatory feedback [8], [9]. This breath-to-breath variability has been modulated with background hyper- and hypocapnia [10] but has not yet been applied to investigation of drug action.

In the present paper we have examined the effect of remifentanil infusion on respiratory control by quantifying the dynamic interrelationships between PETCO₂, V_T and breath-to-breath ventilation (V_T/T_{TOT}, where T_{TOT} is total breath time) in both causal directions of the ventilatory loop. For this purpose, we have used a nonlinear modeling approach (Laguerre expansion technique) [11]. First, we considered the dynamic influence of spontaneous PETCO₂ fluctuations on ventilatory variability (the forward part of the ventilatory loop), whereby the latter was assessed by tidal volume and breath-to-breath ventilation. Subsequently, we examined the influence of ventilatory variability on PETCO₂ (the reverse part of the ventilatory loop) by reversing the roles of input and output. We hypothesized that remifentanil, by depressing chemoreceptor responsiveness, would decrease the influence of PETCO₂ fluctuations on ventilatory variability. On the other hand, we were unsure whether the relationship between ventilatory variability and PETCO₂ variability would remain unaffected.

II. METHODS

A. Subjects and experimental protocol

This study was approved by the Oxfordshire Clinical Ethics research committee. Eleven healthy volunteers (age 27 ± 5 years) were examined. The subjects wore a tight fitting facemask attached to a modified T-piece breathing system. The fresh gas flow rate was 30 litres per minute to eliminate rebreathing of expired gases. Since respiratory depression may cause hypoxemia, which causes activation of peripheral chemoreceptors in the carotid bodies and stimulates respiration, we maintained end-tidal oxygen (PETO₂) tension at 30 kPa. A target-controlled infusion of remifentanyl was delivered via an indwelling intravenous cannula inserted into a vein in the left forearm. Stepwise ascending effect site concentrations of zero (baseline), 0.7, 1.1 and 1.5 ng/ml were maintained with a computer-controlled infusion pump (Graseby 3500 TCI). We chose relatively low doses of remifentanyl to investigate the subtle changes in respiratory control seen at clinically relevant opioid analgesia, as opposed to the much higher doses used in anesthesia.

Oxygen saturations, heart rate, PETCO₂ and PETO₂ were monitored continuously using a Datex Cardiocap II (Datex Instrumentarium), respiratory volume and timing was measured with a turbine respiratory flow meter (VMM-400, Interface Associates). Following a minimum of ten minutes to adapt to the mask, the baseline recordings were taken for 15 minutes, and then for each level of remifentanyl. Five minutes were allowed to reach target effect site concentration; continuous recordings were made for the following 15 minutes at that stable effect site concentration.

B. Mathematical Methods

The general Volterra model for a Q -th order nonlinear system is:

$$y(n) = \sum_{q=0}^Q \sum_{m_1=0}^M \dots \sum_{m_q=0}^M k_q(m_1, \dots, m_q) x(n-m_1) \dots x(n-m_q) \quad (1)$$

where $x(n)$ and $y(n)$ are the system input and output respectively (both PETCO₂ and V_T or V_T/T_{TOT} assume the roles of input and output), M is the system memory and $k_q(m_1, \dots, m_q)$ are the Volterra kernels of the system, which describe the linear ($Q=1$) and nonlinear ($Q>1$) dynamic effects of the input on the output. Eq. (1) reduces to the convolution sum for linear systems, with $k_1(m)$ corresponding to the impulse response of the system.

The impulse response or Volterra kernels can be estimated efficiently from the input-output data, by utilizing function expansions in terms of the orthonormal Laguerre basis:

$$k_q(m_1, \dots, m_q) = \sum_{j_1=0}^L \dots \sum_{j_q=j_{q-1}+1}^L c_{j_1 \dots j_q} b_{j_1}(m_1) \dots b_{j_q}(m_q) \quad (2)$$

where $c_{j_1 \dots j_n}$ are the expansion coefficients, $b_j(m)$ is the j -th order Laguerre function and $L+1$ is the total number of functions that yields an adequate system representation. In matrix form:

$$\mathbf{y} = \mathbf{V}\mathbf{c} + \boldsymbol{\varepsilon} \quad (3)$$

where \mathbf{V} is a matrix that contains the convolution of $x(n)$ with the Laguerre functions $\{b_j\}$ and their second (or higher) order products. The expansion coefficients can be obtained as the least-squares solution of (3) [11]:

$$\mathbf{c}_{\text{est}} = [\mathbf{V}^T \mathbf{V}]^{-1} \mathbf{V}^T \mathbf{y} \quad (4)$$

We used the normalized mean-square error (NMSE) of the output prediction to assess model performance and determine the model complexity (i.e., L and Q) with a statistical criterion [12]. For the chemoreflex pathway (PETCO₂ \rightarrow V, where V corresponds to either V_T or V_T/T_{TOT}) sighs, which were defined as breaths with values greater than 1.5 times the mean breath value were removed by linear interpolation before model estimation, as they are viewed as part of the disturbance component of V (V_d in Fig. 1). Furthermore, a pure time delay of 2 breaths was hypothesized in the effects of PETCO₂, in order to account for the previously described transport delays in the action of PETCO₂. Sighs were not removed for the reverse pathway (V \rightarrow PETCO₂), since they are a significant determinant of PETCO₂ changes. The spectral power (SP) of the first- and second-order kernels was calculated by integrating their discrete-time Fourier transforms from 0 to 0.3 cycles/breath for k_1 and from [0 0] to [0.3 0.3] cycles per breath for k_2 . Changes in the steady-state values and the SP of the respiratory variables, as well as in the SP of the first and second-order kernels were assessed by using repeated measures ANOVA.

III. RESULTS

The effects of remifentanyl on the respiratory variables are shown in Table I. We observed a dose-dependent decrease in respiratory rate that was due to increases in duration of expiratory time (TE). V_T initially decreased but increased at higher levels towards baseline values. PETCO₂ increased and became more variable. The SP of PETCO₂ and V_T between 0 and 0.3 cycles/breath revealed an increase over the entire frequency range for PETCO₂ and a less pronounced increase for V_T above 0.02 cycles/breath.

The prediction NMSEs obtained by linear and nonlinear models ($Q=1, 2$) for both pathways of the ventilatory loop are given in Table II. Nonlinear models improved model performance in both pathways. For the forward part of the loop (PETCO₂ \rightarrow V) we also observed a dose-dependent reduction in the NMSE values during remifentanyl (nonlinear models), which suggests that a larger fraction of the respiratory variability is caused by PETCO₂ changes. Similar results were obtained for V_T/T_{TOT} .

TABLE I
RESPIRATORY PARAMETERS

	Baseline	0.7 ng/ml	1.1 ng/ml	1.5 ng/ml
PETCO ₂ (kPa)	5.4 (0.3)	5.8 (0.4)**	5.9 (0.4)**	6.2 (0.6)**
SP PETCO ₂	1.2 (1.1)	2.6 (1.5)**	3.3 (2.9)	4.0 (2.6)**
PETO ₂ (kPa)	30 (1)	30 (1)	30 (2)	29 (2)
V_T (ml)	412 (134)	348 (128) *	360 (114)	384 (144)
SP V_T	$1.7 \cdot 10^2$ (1.3)	$1.8 \cdot 10^2$ (1.7)	$3.2 \cdot 10^2$ (3.3)	$2.5 \cdot 10^2$ (2.3)
Ti (s)	1.5 (0.3)	1.6 (0.4)	1.6 (0.5)	1.6 (0.5)
TE (s)	2.7 (0.4)	3.8 (0.8)**	4.7 (1.6)**	5.2 (2.1)**
V_T/T_{TOT} (ml/s)	100.2 (23.3)	64.0 (23.6)**	60.8 (20.6)**	61.5 (22.3)**
SP V_T/T_{TOT}	436.0 (233.9)	494.2 (323.7)	658.9 (430.2)	989.4 (1274.5)
HR	56.8 (7.8)	55.8 (7.1)	54.8 (9.3)	55.9 (8.3)

Values are mean (SD). * $P < 0.05$ ** $P < 0.01$ with respect to baseline.

TABLE II
PREDICTION NMSES FOR THE FORWARD AND REVERSE PATHWAYS
OF THE VENTILATORY LOOP

	NMSE [%] Mean (SE)			
	PETCO ₂ →V _T		V _T →PETCO ₂	
Baseline	87.6 (2.0)	71.0 (2.9)	71.1 (5.0)	53.3 (4.6)
0.7 ng/ml	87.3 (3.0)	62.6 (3.6)	67.5 (5.5)	46.6 (4.6)
1.1 ng/ml	89.4 (1.6)	62.1 (3.9)	79.7 (1.6)	58.8 (3.8)
1.5 ng/ml	87.1 (2.5)	57.9 (3.7)	78.6 (4.4)	54.3 (4.5)

Representative data sets used for model estimation (baseline) are shown in Fig. 2, along with the corresponding linear and nonlinear model predictions. PETCO₂ variations mainly account for the V_T post-sigh response, as sighs are clearly correlated with sharp PETCO₂ drops, which in turn influence V_T. These sharp drops are accounted by the V_T → PETCO₂ model. In the frequency domain, the incorporation of nonlinear model terms improved performance over a wide range of frequencies below 0.03 cycles/breath.

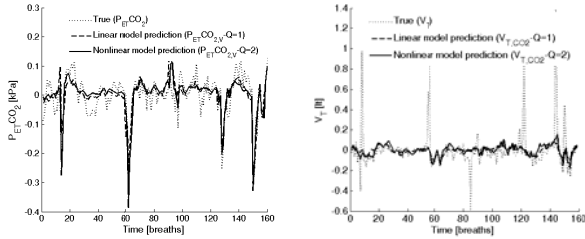


Fig. 2. PETCO₂ and V_T time series (dotted lines) during baseline, used for model estimation in both pathways of the ventilatory loop, and corresponding model predictions (PETCO_{2,V} and V_{T,CO2} respectively, solid lines). The large drops in PETCO₂ induced by sighs are clearly accounted by the V_T → PETCO₂ model (left), while PETCO₂ changes account mainly for the post-sigh V_T response (right).

The averaged impulse responses ($Q=1$ in (1)) for the forward part of the ventilatory loop are displayed in Fig. 3, when both V_T (blue) and V_T/T_{TOT} (black) were used to assess ventilatory variability. The form of the averaged impulse response during baseline suggests that an increase in PETCO₂ will cause an increase in V_T (or V_T/T_{TOT}), with the maximum instantaneous effects occurring at 4 and 8 breaths after the PETCO₂ increase. The impulse response values generally decreased during remifentanyl infusion, with the decrease being more evident for the second peak.

The impulse responses of Fig. 3 in the frequency domain during baseline exhibit a main resonant peak occurring between 0.04 and 0.08 cycles/ breath. These characteristics were less consistent during remifentanyl infusion, with the main resonant peak being shifted to lower frequencies. The averaged impulse response for the reverse branch of the ventilatory loop (V → PETCO₂) is shown in Fig. 4 for both V_T (blue) and V_T/T_{TOT} (black). Its form suggests that an increase in ventilation will lead to a decrease in PETCO₂, with the maximum effects occurring within the first 2 breaths. The dynamic effects of these changes cease before 20 breaths. Remifentanyl did not alter these characteristics; however, the impulse response values increased at all levels, suggesting a stronger dynamic effect of ventilatory variability on PETCO₂.

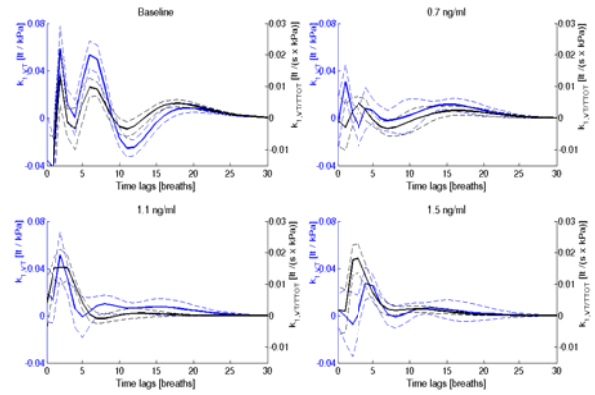


Fig. 3. Averaged impulse response for the chemoreflex pathway, whereby both V_T (blue) and V_T/T_{TOT} (black) were used to assess ventilation. Remifentanyl decreased the impulse response values.

The above observations are quantified by the SP of the first- and second-order kernels for both pathways shown in Fig. 5. For the chemoreflex branch (left), a decrease was observed in the SP of k_1 and k_2 . This decrease was statistically significant only during the lowest level of remifentanyl infusion for V_T; however, more pronounced differences were observed when V_T/T_{TOT} was used as the model output. For the reverse pathway (right), the SP of k_1 increased significantly ($P<0.01$ during all remifentanyl levels for V_T). The SP of the second-order model components k_2 increased as well ($P<0.05$ at 0.7 ng/ml).

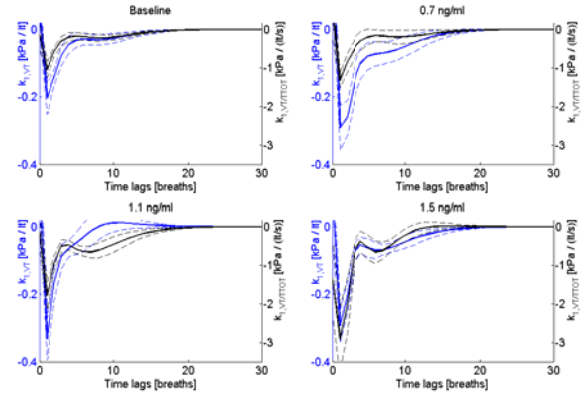


Fig. 4. Averaged impulse response for the V → PETCO₂ pathway when both V_T (blue) and V_T/T_{TOT} (black) were used as measures of ventilatory variability. A main negative peak is observed at 1 breath. The impulse response drops to zero before 20 breaths in all cases. The impulse response values increased considerably at all infusion levels.

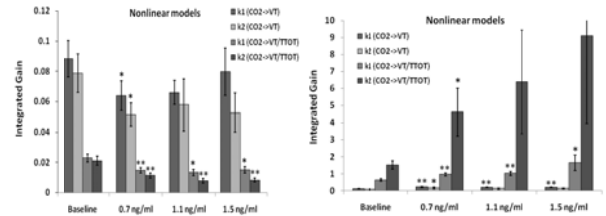


Fig. 5. Spectral power of the first and second-order Volterra kernels for the chemoreflex (left) and reverse (right) pathways. The spectral power of both the linear and nonlinear chemoreflex components between 0 and 0.3 cycles/ breath decreased during remifentanyl infusion. * $P<0.05$, ** $P<0.01$ compared to baseline.

IV. DISCUSSION

We have demonstrated that infusion of remifentanyl leads to increased irregularity of the respiratory pattern during spontaneous respiration. Moreover, it decreased the strength of the dynamic effect of natural PETCO₂ variability on tidal volume and breath-to-breath ventilation, but increased the reverse relationship, i.e., the effect of ventilatory variability on PETCO₂. Nonlinear, rather than linear, models best described these dynamic relationships. Collectively, these findings suggest the potential use of data-driven approaches to identify drug-induced changes on respiratory control on a systems level, employing minimally invasive protocols.

The influence of the spontaneous variability of PETCO₂ on fluctuations of breath-to-breath V_T variability has been demonstrated by application of a PETCO₂ buffering technique [7]. Moreover, coherent oscillations between PETCO₂ and mean inspiratory flow V_I/T_I in the frequency domain, in agreement with a closed-loop model of the chemoreflex feedback, have been reported [9]. In order to quantify these correlations, we have employed a nonlinear approach that extends the aforementioned observations to both pathways of the ventilatory loop.

The high NMSE values for both pathways reflect the fact that the resting variability of PETCO₂ and V is determined by many factors. Significant influences that are part of the disturbance signal (non-chemical respiratory drive) V_d are exerted by cortical centres (behavioural, volitional modulation), the reticular activating system during wakefulness, as well as changes in cardiovascular parameters (e.g. cardiac output, cerebral blood flow oscillations) [13]. Our results (nonlinear models) suggest an increase in the chemical drive of ventilation during remifentanyl infusion. Since the magnitude of the PETCO₂ → V model dynamics decreased, this increase was due to the increased PETCO₂ variability. Likewise, the PETCO₂ disturbance signal is influenced by the aforementioned and other factors, such as metabolism, level of arousal, sleep state and PaO₂ level [14].

The presence of nonlinear dynamics in spontaneous respiratory volume variability has been reported [15]. The steady-state chemoreflex response, i.e. the relation between the mean values of PETCO₂ and ventilation has been shown to be relatively linear; however, in the present study we are examining the dynamic relation between spontaneous fluctuations around the mean values. Regarding the reverse branch, it has been suggested that the influence of V_T on CO₂ exhibits nonlinear characteristics in the case of large V_T variations [9]. The Laguerre expansion technique was used in [16] to obtain the loop gain in normal subjects and patients with obstructive sleep apnea. However, the ventilatory impulse response was estimated as a whole, whereas we have examined the two branches separately.

The form of the impulse response between PETCO₂ and V agrees qualitatively with the previously described effects of PETCO₂ on ventilation, i.e., an increase in PETCO₂ results in an increase in ventilation some breaths later. The spectral peaks of the PETCO₂ → V dynamic models during baseline agree with the results of [9], where coherent oscillations

between PETCO₂ and V_I/T_I below 0.15 cycles/breath with a peak at 0.08 cycles/breath were reported. The multi-phasic characteristic observed during baseline possibly reflects the closed loop nature of respiratory control, as an initial perturbation in PETCO₂ will cause a change in tidal volume that has an opposite effect on PETCO₂. The spectral power of the PETCO₂ → V system dynamics decreased during remifentanyl administration, suggesting decreased chemosensitivity. We speculate that the increased PETCO₂ variability seen in the present study may help maintain ventilation during opioid-induced respiratory depression.

The waveforms of the V → PETCO₂ impulse responses correspond to the well-known effects of respiratory changes, i.e., an abrupt increase in V_T (or V_T/T_{TOT}) results in a rapid decrease in PETCO₂. Remifentanyl infusion did not have a profound effect upon the timing of the V → PETCO₂ impulse responses; however, its increased values suggest increased PETCO₂ sensitivity to ventilatory changes. Hypercapnia may contribute to this increased, due to greater CO₂ excretion per breath as a simple mass effect, as similar observations have been reported during sleep, which also induces hypercapnia.

REFERENCES

- [1] K.T. Pattinson "Opioids and the control of respiration," *Br J Anaesth* 100: 747-758, 2008.
- [2] N.M. Mellen, W.A. Janczewski, C.M. Bocchiaro and J.L. Feldman "Opioid-induced quantal slowing reveals dual networks for respiratory rhythm generation," *Neuron* 37: 821-826, 2003.
- [3] T. Bouillon, J. Bruhn, H. Roepcke and A. Hoeft "Opioid-induced respiratory depression is associated with increased tidal volume variability," *Eur J Anaesth* 20: 127-133, 2003.
- [4] M.E. Pedersen, M. Fatemian and P.A. Robbins "Identification of fast and slow ventilatory responses to carbon dioxide under hypoxic and hyperoxic conditions in humans," *J Physiol* 521: 273-287, 1999.
- [5] P.L. Bailey, J.K. Lu, N.L. Pace, J.A. Orr, J.L. White, E.A. Hamber, M.H. Slawson, D.J. Crouch and D.E. Rollins "Effects of Intrathecal Morphine on the Ventilatory Response to Hypoxia," *The New England Journal of Medicine* 343: 1228-1234, 2000.
- [6] A. Dahan, A. Yassen, H. Bijl, R. Romberg, E. Sarton, L. Teppema, E. Olofsen and M. Danhof "Comparison of the respiratory effects of intravenous buprenorphine and fentanyl in humans and rats," *Br J Anaesth* 94: 825-834, 2005.
- [7] M. Modarreszadeh and E.N. Bruce "Ventilatory variability induced by spontaneous variations of PaCO₂ in humans," *J Appl Physiol* 76: 2765-2775, 1994.
- [8] M.C.K. Khoo and V.Z. Marmarelis "Estimation of peripheral chemoreflex gain from spontaneous sigh responses," *Ann Biomed Engin* 17: 557-570, 1989.
- [9] J.G. van den Aardweg and J.M. Karemaker "Influence of chemoreflexes on respiratory variability in healthy subjects," *Am J Respir Crit Care Med* 165: 1041-1047, 2002.
- [10] B.F. BuSha and M.H. Stella "State and chemical drive modulate respiratory variability," *J Appl Physiol* 93: 685-696, 2002.
- [11] V.Z. Marmarelis "Identification of nonlinear biological systems using Laguerre expansions of kernels," *Ann Biomed Eng* 21: 573-589, 1993.
- [12] J. Sjöberg "Non-linear System Identification With Neural Networks," Linköping, Sweden: Linköping University, 1995.
- [13] M.C.K. Khoo "Determinants of ventilatory instability and variability," *Resp. Physiol.* 122: 167-182, 2000.
- [14] A. Crosby P.A. and Robbins "Variability in end-tidal PCO₂ and blood gas values in humans," *Exp. Physiol.* 88: 603-610, 2003.
- [15] M. Wysocki, M.N. Fiamma, C. Straus, C.S. Poon and T. Similowski "Chaotic dynamics of resting ventilatory flow in humans assessed through noise titration," *Respir Physiol Neurobiol* 153: 54-65, 2006.
- [16] M.H. Asyali, R.B. Berry and M.C.K. Khoo "Assessment of closed-loop ventilatory stability in obstructive sleep apnea," *IEEE Trans Biomed Engin* 49: 206-216, 2002.